mtDNA Sequence Data The Phylogenetic Position of Megachasma pelagios Inferred from

John F. Morrissey¹, Katherinc A. Dunn², and Francesco Mulé

Keywords: mtDNA, phylogeny, megamouth, Megachasma, Lamniformes

Cetorhinus, suggesting independent evolution of filter feeding within order Lamniformes that Megachasma is a) the most primitive living lamniform shark and b) not the sister-group of was not affected by a variety of weighing schemes nor by deletion of outgroup taxa. parsimonious tree with a length of 104 steps and a consistency index of 0.779. The topology of this tree sequenced, and analyzed phylogenetically using parsimony. These analyses yielded a single mostlamniform genera and three outgroup taxa. A portion of the 12S rRNA mitochondrial gene was amplified. results have been contradictory. The objective of this study was to infer the phylogenetic position of Megachasma within order Lamniformes using molecular data. DNA was extracted from seven of ten Studies on the relationship of Megachasma to other lamnoids have been infrequent and The results suggest

beyond the scope" of their paper. extant lamnoid sharks, but stated that "a detailed account of lamnoid interrelationships is Finally, they tentatively suggested that Megachasma may be the primitive sister-group of all Megachasma from all other lamniform families, including the other filter feeder, Cetorhinus. al. (1983) listed an impressive array of phenetic characters as evidence for the separation of musculature, and spiral valve intestine as evidence of its lamnoid affinity. Moreover, Taylor et mouth, eyes, teeth, claspers, dorsal fin skeleton, chondrocranium, pectoral fin skeleton, jaw chondrocranium, and vertebral column. They went on to list phenetic characters of its nostrils, phenetic characters of its head, spiracles, external gill slits, placoid scales, girdles, fins, Compagno, 1973, 1977). Taylor et al. (1983) stated that its galcomorph affinity is revealed by elasmobranch known to be a deep-sca planktivore (Diamond, 1985). Its formal description (Taylor et al., 1983) suggested that Megachasma is a lamniform galeomorph shark (sensu Mobula, and Rhincodon, are scattered among three elasmobranch orders), and the first represents only the fifth genus of filter-feeding elasmobranch (the others, Cetorhinus, Manta, cm) should not have allowed this species to remain undiscovered for so long. Second, it naval research vessel AFB-14. Its discovery was surprising for two reasons. First, its size (446 The first specimen of Megachasma pelagios was captured on 15 November 1976 by the

speculated that his Cetorhinidae (i.e. Cetorhinus + Megachasma) may be the sister-group of and Megachasma form a confamilial group of lamniforms specialized for filter feeding. He Cetorhinus and Megachasma tallied by Taylor et. al (1983), but still suggested that Cetorhinus Megachasma. The first was Maisey (1985), which acknowledged the many differences between particular, and lamnoid interrelationships, in general, since the formal description of Two studies have broached the question of megamouth's phylogenetic position, in

^{**}Department of Biology, 114 Hofstra University, Hempstead, NY 11550-1090, USA ***Department of Wildlife and Fisheries Scionces, Texas A & M University, College Station, TX 77803-2258, USA

Phylogenetic Position of Megamouth -35

other advanced lamnoid families that possess plesodic pectoral skeletons (i.e., Lamnidae and Alopiidae), and that *Megachasma* is not the primitive sister to all living lamnoid sharks. Implicit in Maisey's (1985) phylogenetic hypothesis is the contention that the filter-feeding mechanism of *Megachasma* and *Cetorhinus* is autapomorphous for his family Cetorhinidae. Hence, filter feeding evolved only once among the lamnoids.

Compagno (1990) rejected Maisey's (1985) confamilial classification of Cetorhinus and Megachasma by summarizing a great deal of phenetic evidence of their morphological dissimilarity and by contradicting the five sets of synapomorphies of Maisey's (1985) Cetorhinidae. Compagno (1990) went on to present a cladistic analysis of all living lamnoid sharks. His phylogenetic hypothesis suggested that a) Megachasma is not the primitive sistergroup of all other lamniforms, b) Megachasma + Cetorhinus do not form a monophyletic group (and acquired their filtering mechanisms independently), and c) lamniforms with plesodic pectoral skeletons (i.e., Megachasmidae, Alopiidae, Cetorhinidae, and Lamnidae) are most derived. Nevertheless, Compagno (1990) admitted that "there are problems with the cladogram...and supporting arguments." Therefore, the present study was undertaken to infer the phylogenetic position of Megachasma within the order Lamniformes using molecular data.

Materials and Methods

Skeletal muscle samples were obtained from the following seven lamniform species (which represent seven of ten extant genera): Alopias vulpinus, Carcharodon carcharias, Cetorhinus maximus, Isurus oxyrinchus, Lamna nasus, Megachasma pelagios, and Odontaspis ferox. Skeletal muscle samples also were obtained from three outgroup genera (Heptranchias perlo, Hydrolagus colliei, and Ginglymostoma cirratum). The skeletal muscle was digested and the DNA was extracted following Dunn and Morrissey (1995).

A portion of the 12S rRNA mitochondrial gene was amplified with the primers of Kocher et al. (1989) via the polymerase chain reaction (PCR). Successful amplification was confirmed by electrophoresis of the PCR product on a 1.4% agarose gel and staining with ethidium bromide. Excess primers were removed enzymatically, the PCR product was sequenced manually (Sequenase version 2.0; US Biochemical), and the reaction was visualized on a 6% acrylamide gel (Dunn and Morrissey, 1995).

The resulting nucleotide sequences were edited with SeqApp (version 1.9al69; Gilbert, 1992), aligned with Clustal (version V; Higgins et al., 1992), and analyzed with Phylogenetic Analysis Using Parsimony (PAUP; version 3.1; Swofford, 1990). All topologies under a variety of weighting schemes were examined using the EXHAUSTIVE SEARCH option with ACCTRAN optimization of the unordered (FITCH) characters. Confidence in the resulting topology was estimated using the BOOTSTRAP (Felsenstein, 1985) and BRANCH AND BOUND (Hendy and Penny, 1982) options of PAUP.

Results

The nucleotide sequences of a homologous 203-base-pair region of the 12S rRNA mitochondrial gene were obtained for the seven ingroup and three outgroup taxa. By weighting transitions and transversions equally, a single most-parsimonious tree was obtained (Fig. I) with a length of 104 steps and a consistency index (Cl; Kluge and Farris, 1969) of 0.779. Eight one-step-off trees were found and the distribution of lengths of all possible tree topologies was skewed significantly (gl = -0.98; t = 6.43; P < 0.01) to the left. Next, the possibility of saturation of transitions, a common occurrence in sharks (Martin, 1995), was evaluated by weighting transversions two and ten times more than transitions. A tree identical to Figure 1

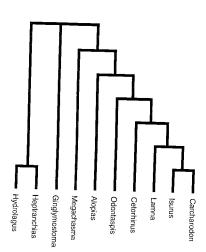


Fig. 1. Single most-parsimonious tree obtained using the EXHAUSTIVE SEARCH option and ACCTRAN optimization in PAUP. Length = 104 steps, CI=0.779.

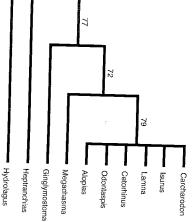


Fig. 2. Majority rule (50%) consensus tree derived from 1000 bootstrap replications. Values at each node represent the percentage of bootstrap replicates possessing that node.

was obtained in both cases. Moreover, an identical resolution of the ingroup taxa was obtained using only *Heptranchias* as the outgroup (65 steps; CI = 0.831), only *Hydrolagus* as the outgroup (75 steps; CI = 0.867), or by excluding *Ginlymastoma* from the analysis (87 steps; CI = 0.828). One thousand replications of a 50% majority-rule bootstrap analysis resulted in Figure 2. All fully resolved nodes were found in at least 72% of the trees (i.e., there is a probability of ≥ 95% that those clades are real; Hillis and Bull, 1993).

Discussion

The phylogenetic position of Megachasma has been considered infrequently. Only two

attacked the suggestion that megamouth and basking sharks are confamilial, and suggested that Megachasma is relatively derived and sister to a clade containing Alopiidae, Cetorhinus, and Compagno (1990) disagreed with the idea that Megachasma is the most primitive lamnoid suggested that megamouth sharks are rather derived (based on their possession of a plesodic pectoral fin skelcton) and form a monophyletic family with basking sharks, Cetorhinus. published since the discovery of the species in 1976 (Taylor et al., 1983). Maisey (1985) phylogenetic hypotheses regarding the relationship of megamouth to other lamnoids have been

in many Paleozoic elasmobranchs (Compagno, 1973; Zangerl, 1973; Bendix-Almgreen, 1975). to be primitive to all living lamniform sharks. Perhaps they were skeptical of the polarity of this character due to its homoplasious distribution among extant sharks and its well-known presence megamouth possessed a plesodic skeleton in its pectoral fin, yet still considered Megachasma and sphyrnids) (Compagno, 1988). However, Taylor et al. (1983) clearly stated that Alopias, Cetorhinus, and all lamnids), and some carcharhiniforms (hemigaleids, carcharhinids, (Stegostoma, Ginglymostoma, Nebrius, and Rhincodon), some lamniforms (Megachasma, far into the fin to augment the ceratotrichia) are variously displayed by some orectolobiforms plesodic skeleton in its pectoral fins. Plesodic pectoral fins (wherein the pectoral radials extend most primitive lamnoid was prevented by its possession of a supposedly derived character, a palatoquadrate). Recognition by Maisey (1985) and Compagno (1990) of Megachasma as the originally suggested by Taylor et al. [1983] on the basis of primitive characters of the teeth and suggests that Megachasma is, in fact, the primitive sister-group of all living lamniforms (as confusion surrounding the relationships of Megachasma to other lamnoid sharks. Figure 1 The phylogenetic hypothesis presented in this study (Fig. 1) may clear up some of the

Miyamoto and Fitch, 1995) before the interrelationships of lamniform taxa can be resolved Carcharodon. Clearly, the accumulation of additional molecular data is necessary (Hillis, 1995) the most derived lamniform family, and c) that Lamna is the sister-group to Isurus + Specifically, it confirms the idea a) that lamnid sharks form a single clade, b) that lamnids are well-documented relationships within order Lamniformes (see Compagno, 1977, 1990) within a data set (Huelsenbeck and Hillis, 1993). Finally, Figure 1 also supports several other fully with confidence internodes (i.e., rapid speciation) has been theorized to camouflage the phylogenetic signal with rapid speciation events, phenomena which are manifested in the lack of resolution of these Cappetta, 1987; Ward and Wiest, 1990, Siverson, 1992, 1995) must have occurred concurrently separated from all other lamnoid families, is not unique to the present study (see Taylor et al., lamnoid species. This combination of long branch lengths (i.e., ancient lineages) and short other lamnoids. This notion, that Megachasma represents a very distinct taxon that is well analysis (Fig. 2), two things are clear. First, Megachasma is primitive to and distinct from all 1983; Compagno, 1990). Second, the ancient appearance of many lamnoid lineages (e.g., see Although the interrelationships within the lamnoids were made dubious by a bootstrap

Acknowledgments

symposium volume. We thank D.E. Pumo for advice and laboratory equipment. We thank D nakamichi for providing a muscle sample from Megachasma pelagios #7, the subject of this Fournier, K. Goldman, G. Naylor, and G. Van Dykhuizen for kindly providing tissue samples from Hofstra University to JFM. Burke, R. Moritz and M. O'Keefe for logistical support. This research was funded by grants from lamniform fishes. We especially thank K. Yano and the staff of Marine World umino-We thank C. Baduini, the Bayshore Mako Tournament, J. Castro, G. Croft, J. Ellis, R

Literature Cited

Bendix-Almgreen, S.E. 1975. The paired fins and shoulder girdle in *Cladoselache*, their morphology and phyletic significance. Colloques Int. Cent. Nam. Rech. Sci., Paus, 218. 111-123. Capetta, H. 1987. Chondrichthyes II. Mesozoic and Cenozoic Elasmobranchii. Handbook of Paleoichthyology, Vol. 3.B. Gustav Fischer Verlag, Stuttfart, iii + 193 pp.

Compagno, L.J.V. 1973. Interrelationships of living elasmobranchs. Pages 15-61 in P.H. Greenwood, R.S. Miles, and C. Patterson, eds. Interrelationships of fishes. Zool. J. Linn. Soc., 53. Suppl. 1. Compagno, L.J.V. 1977. Phyletic relationships of living sharks and rays. Amer. Zool., 17: 303-322. Compagno, L.J.V. 1988. Sharks of the order Carcharhiniformes. Princeton University Press, Princeton. 486 pp.,

Compagno, I.J.V. 1990. Relationships of the megamouth shark, *Megachasmia pelagios* (Lamniformes: Megachasmidae), with comments on its feeding habits. Pages 357-379 in H.L. Pratt, Ir., S.H. Gruber, and T. Tanuchi, eds. Elasmobranchs as living resources: advances in the biology, ecology, systematics, and the status of fisherics. NOAA Tech. Rep. NMTS, 90.

Diamond, J.M. 1985. Filter-feeding on a grand scale. Nature 316: 679-680.

Dunn, K.A. and J.F. Morrissey. 1995. Molecular phylogeny of clasmobranchs. Copeia, 1995 (3): 526-531.

Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using bootstrap. Evolution, 39: 783-791. Gilbert, D. G. 1992. A biological sequence editor and analysis progrom for Macinnosh computers that can be obtained thru anonymous fip to fip-bio.indiana.edu, in folder /molbio/seqapp, as seqapp.hqx. Hendy, M.D. and D. Penmy. 1982. Branch and bound algorithms to determine minimal evolutionary trees. Math. Biosci., 59: 277-290.

Higgins, D.G., A.J. Bleasby, and R. Fuchs, 1992. Clustal V: Improved software for multiple sequence

Huelsenbeck, J.P. and D.M. Hillis. 1993. Success of phylogenetic methods in the four-taxon case. Syst. Biol. alignment. Comput. Appl. Biosci., 8: 189-191.

Hillis, D.M. 1995. Approaches for assessing phylogenetic accuracy. Syst. Biol., 44: 3-18.

Hillis, D.M. and J.J. Bull. 1993. An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. Syst. Biol., 42: 182-192.

Kluge, A.G. and J.S. Farris, 1969. Quantitative phyletics and the evolution of anurans. Syst. Zool., 18: 1-32. Kocher, T.D., W.K. Thomas, A. Meyer, S.V. Edwards, S. Paabo, F.X. Villablanca, and A.C. Wilson, 1989.

Dynamics of mitochondrial DNA evolution in animals: amplification and sequencing with conserved primers. Proc. Natl. Acad. Sci., 86: 6196-6200.

Martin, A.P. 1985. Relationships of the megamouth shark, Megachasma. Copeia, 1985(1): 228-231.

Martin, A.P. 1995. Mitochondrial DNA sequence evolution in sharks: rates, patterns, and phylogenetic inferences. Mol. Biol. Evol., 12: 1114-1123.

Miyamoto, M.M. and W.M. Fitch. 1995. Testing species phylogenies and phylogenetic methods with congruence. Syst. Biol., 44: 64-76.

Siverson, M. 1992. Biology, dental morphology, and taxonomy of lamniform sharks from the Campanian of the Kristianstad Basin, Sweden, Paleontology, 35: 519-554.

Siverson, M. 1995. Revision of the Danian cow sharks, sand tiger sharks, and goblin sharks (Hexanchidae, Odontaspidae, and Mitsukurinidae) from southern Sweden. J. Vert. Palacontol., 15: 1-12. Swofford, D.L. 1990. Phylogenetic analysis using parsimony (PAUP), version 3.1. Illinois Natural History

Survey, Urbana.

Survey, Urbana.

Taylor, L.R., I.J.V. Compagno, and P.J. Struhsaker, 1983. Megamouth - a new species, genus, and family of lamnoid shark (Megachasma pelugios, family Megachasmidae) from the Hawaiian Islands. Proc. Calif. Acad. Sct., 43(8): 87-110.

Ward, D.J. and R.L. Wiest, Jr. 1990. A checklist of Paleocene and Eocene sharks and rays (Chondrichthyes) for the Pannunkey Group, Maryland and Virginia, USA. Tertiary Res., 12: 81-88.

Zangerl, R. 1973. Interrelationships of early chondrichthyans. Pages 1-14 in P.H. Greenwood, R.S. Miles, and C. Patterson, eds. Interrelationships of fishes. Zool. J. Linn. Soc., 53, Suppl. 1.